

LITTOISTENJÄRVEN seuranta sinilevämyrkkyjen suhteen

Date of analysis: 11.9.2020

Sample collection, immunoassay, data analysis and report by SULTANA AKTER

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Assay method reference:

Sultana Akter, Markus Vehnäläinen, Lisa Spoof, Sonja Nybom, Jussi Meriluoto, and Urpo Lamminmäki. Broad-spectrum noncompetitive immunocomplex immunoassay for cyanobacterial peptide hepatotoxins (microcystins and nodularins). Analytical Chemistry, 2016, 88, 10080–10087. (PMID:27657987)

Assay method: Immunoassay based on Akter et al., 2016 with slight modification

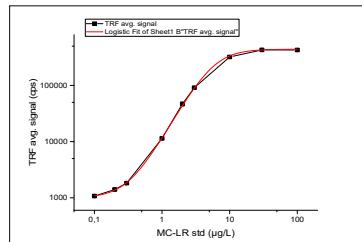
- Prewash streptavidin coated strips (yellow, normal, Lot KG1739).
- Add blank (reagent water), MC-LR standard or sample, 50 µL/well as Triplicate.
- Add Reagent Mixture, 50 µL/well
- Incubate with slow shaking for 1 hour at RT.
- Wash 4 x.
- Add Enhancement solution 200 µL per well. Use the Plate Dispenser.
- Incubate with slow shaking for 10 min at RT.
- Measure the Time resolved fluorescence (TRF) signal with Plate fluorometer.
- Resolve standard curve with Origin 2016 and logistic fit.

microcystin-LR (MC-LR) standard

MC-LR (Enzo Life sciences, ALX350-431)

Prepared original stock of 1000 µg/L in reagent water+5%Methanol. Stored at (-20°C)

30.9.2019SA: Further working standard solution in reagent water: 100, 30, 10, 3, 2, 1, 0.3, 0.2 and 0.1 µg/L



standard curve of microcystin-LR

Reagent mixture in assay buffer

1 µg/mL biotinylated anti-ADDA Antibody (stock 256 µg/ml); +

1 µg/mL anti-immunocomplex scFv-AP (stock 440 µg/ml); +

0.5 µg/ml N1-Eu-anti AP pAb (stock 200 µg/ml, 16.1.2020);

(x)	TRF signal (counts per second)			(y)			
MC-LR (µg/L) std	A	B	C	avg sig	std dev	cv%	blk+3SD (n=9)
0	988	1054	1076	1006	86	8.5	1263
0	1114	911	944				
0	1102	987	874				
0.1	1131	1046	1045	1074	49	4.6	
0.2	1474	1376	1362	1404	61	4.3	
0.3	1890	1714	1863	1822	95	5.2	
1	11666	11598	10961	11408	389	3.4	
2	50216	46893	43408	46839	3404	7.3	
3	93409	89452	90143	91001	2114	2.3	
10	328484	314136	318427	320349	7365	2.3	
30	427007	431886	428430	429108	2509	0.6	
100	410765	455097	416800	427554	24043	5.6	

sample	TRF signal			sig comments	std dev	cv%	*(x) From origin	DF	1x conc (µg/L)	reported conc (µg/L)
	A	B	C	Avg						
A_Saarten taus	1_A	1346	1234	1136	1239	8.5	0.16	1	0.16	<0.2
B_Koilliselkä	2_B	1226	1002	1334	1187	14.3	0.15	1	0.15	<0.2
C_Luoteisselkä	3_C	1212	1246	1131	1196	4.9	0.15	1	0.15	<0.2
A'_Hiekkaranta	4_A'	1156	1214	1306	1225	6.2	0.16	1	0.16	<0.2
B'_Pirttan laituri(1), near Littoistenjärvi	5_B'	1204	1276	1096	1192	7.6	0.15	1	0.15	<0.2
C'_Bussilaituri(2) Rauhaniemi, bus stop 6_C'	6_C'	1144	1210	1162	1172	2.9	0.14	1	0.14	<0.2
D'_Ristikallion Uimaranta	7_D'	1269	1214	1111	1198	6.7	0.15	1	0.15	<0.2
E'_Kuoviuloto	8_E'	1142	1138	1266	1182	6.2	0.14	1	0.14	<0.2
F'_Rantapolun laituri(3)	9_F'	1343	1404	1190	1312	below blk+3SD	110	8.4	0.18	0.18

DL based on (blk+3SD) sig	1263	0.17	µg/L
DL based on true standard above (blk+3SD) signal	1404	0.2	µg/L

Interpretation (11.9.2020 SA)

Raw water samples were analyzed fresh on 11.9.2020.

Before analysis, samples were heated at 80 °C for 10 min to release cell bound toxins if any.

Hence, the results represent the total peptide hepatotoxin amount (already released toxin in water and the cell bound toxin).

The immunoassay (Akter et al., 2016) detects cyanobacterial peptide hepatotoxins (eg microcystins).

For quantification, microcystin-LR was used as standard.

Result:

In Littoistenjärvi water sample of 11.9.2020, the detected cyanobacterial peptide hepatotoxin concentrations (µg/L) (free and cell bound) were shown below from the following samples:

A_Saarten taus: <0.2 µg/L
B_Koilliselkä: <0.2 µg/L
C_Luoteisselkä:<0.2 µg/L

A'_Hiekkaranta:<0.2 µg/L
B'_Pirttan laituri, near Littoistenjärvi: <0.2 µg/L
C'_Bussilaituri, Rauhaniemi, bus stop 6378 : <0.2 µg/L

D'_Ristikallion Uimaranta: <0.2 µg/L
E'_Kuoviuloto: <0.2 µg/L
F'_Rantapolun laituri:<0.2 µg/L

