

LITTOISTENJÄRVEN seuranta sinilevämyrkköjen suhteen

Date of analys 3.8.2022

Sample collection, immunoassay, data analysis and report by SULTANA AKTER

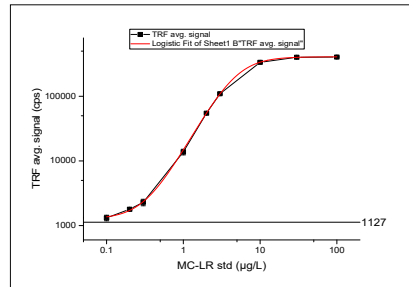
Sultana Akter (sultana.akter@utu.fi), Project Researcher, Department of Life Technologies (Biotechnology), Faculty of Technology, University of Turku

Assay method reference:

Sultana Akter, Markus Vehniäinen, Lisa Spoo, Sonja Nybom, Jussi Meriluoto, and Urpo Lamminmäki. Broad-spectrum noncompetitive immunocomplex immunoassay for cyanobacterial peptide hepatotoxins (microcystins and nodularins), Analytical Chemistry, 2016, 88, 10080–10087. (PMID:27657987)

Assay method: Immunoassay with some modification based on concept of Akter et al., 2016

1. Prewash streptavidin coated strips (yellow,41-07 TV, KG 2109).
2. Add blank (reagent water), MC-LR standard or sample, 50 µL/well as Triplicate.
3. Add Reagent Mixture, 50 µL/well
4. Incubate with slow shaking for 1 hour at RT.
5. Wash 4 x.
6. Add Enhancement solution 200 µL per well. Use the Plate Dispenser.
7. Incubate with slow shaking for 10 min at RT.
8. Measure the Time resolved fluorescence (TRF) signal with Plate fluorometer.
9. Resolve standard curve with Origin 2016 and logistic fit.



standard curve of microcystin-LR

microcystin-LR (MC-LR) standard

MC-LR (Enzo Life sciences, ALX350-431)

Prepared original stock of 1000 µg/L in reagent water+5%Methanol. Stored at (-20C)

9.6.2022: working standard solution in reagent water: 100, 30, 10, 3, 2, 1, 0.3, 0.2 and 0.1 µg/L

Reagent mixture in assay buffer

1 µg/mL biotinylated anti-ADDA Antibody (stock 242 µg/ml); +

1 µg/mL anti-immunocomplex scFv-AP (stock 440 µg/ml) +

0.5 µg/mL N1-Eu-anti AP pAb (stock 200 µg/ml, 16.1.2020).

(x)	TRF signal (counts per second)			(y)				
MC-LR (µg/L)	A	B	C	avg sig	std dev	cv%	blk+3SD (9 blank)	
0	1186	1126	1144				1627	
0	1284	1358	1332	1210		139		
0	1396	1056	1004					
0.1	1466	1228	1258	1317		130		9.8
0.2	1874	1672	1828	1791		106		5.9
0.3	2344	2006	2521	2290		262		11.4
1	12477	13685	15190	13784		1359		9.9
2	53984	54944	55441	54790		741		1.4
3	106491	109582	112371	109481		2941		2.7
10	334971	340421	331328	335573		4576		1.4
30	404629	397455	404559	402214		4122		1.0
100	397982	413268	402032	404427		7920		2.0

sample	TRF signal			(y)	sig comments	std dev	cv%	*(x) From origin		1x conc (µg/L)	Reported conc (µg/L)
	A	B	C	Avg				conc µg/L	DF		
1_A_Saarten taus	1385	1556	1470	1470	below analytical DL	86	5.8	0.14	1	0.14	below 0.2 µg/L (below analytical DL)
2_B_Koilliselkä	1674	1232	1280	1395	below analytical DL	243	17.4	0.11	1	0.11	below 0.2 µg/L (below analytical DL)
3_C_Luoteisselkä	1354	1224	1450	1343	below analytical DL	113	8.4	--	1	#VALUE!	below 0.2 µg/L (below analytical DL)
4_A'_Hiekkaranta	1248	1249	1349	1282	below analytical DL	58	4.5	--	1	#VALUE!	below 0.2 µg/L (below analytical DL)
5_D'_Ristikallion Uimaranta	1652	1404	1400	1485	below analytical DL	144	9.7	0.14	1	0.14	below 0.2 µg/L (below analytical DL)
blk+3SD (n=9)				1627							

Analytical DL (Detection limit) based on (blk+3SD) sig	1627	0.18 µg/L
set Detection Limit (based on used std signal) for reporting	1791	0.20 µg/L

Interpretation (3.8.2022 SA)

Collection of Raw water samples : 3.8.2022

Immunoassay analysis: 3.8.2022.

Before analysis, samples were heated at 80 °C for 10 min to release cell bound toxins if any.

The results represent the total cyclic peptide hepatotoxin amount (already released toxin in water and the cell bound toxin).

The immunoassay detects cyanobacterial peptide hepatotoxins (microcystins and/or nodularin).

For quantification, microcystin-LR was used as standard.

Result:

In Littoistenjärvi water, the detected cyanobacterial peptide hepatotoxin (free and cell bound microcystin) concentrations (µg/L) are as follows:

- 3.8.2022
- A_Saarten taus: below 0.2 µg/L (below analytical DL)
 - B_Koilliselkä: below 0.2 µg/L (below analytical DL)
 - C_Luoteisselkä: below 0.2 µg/L (below analytical DL)
 - A'_Hiekkaranta: below 0.2 µg/L (below analytical DL)
 - D'_Ristikallion Uimaranta: below 0.2 µg/L (below analytical DL)

