

LITTOISTENJÄRVEN seuranta sinilevämyrkyjen suhteen

Menetelmä: Immunomääritys (Akter et al 2016 / Turun yliopisto)

PVM: 15.8.2019

1. Prewash streptavidin coated strips (yellow, normal, Lot KG1574).
2. Add blank (reagent water), MC-LR standard or sample, 50 µL/well as Triplicate.
3. Add Reagent Mixture, 50 µL/well
4. Incubate with slow shaking for 1 hour at RT.
5. Wash 4 x.
6. Add Enhancement solution 200 µL per well. Use the Plate Dispenser.
7. Incubate with slow shaking for at least 5 min at RT.
8. Measure the Time resolved fluorescence (TRF) signal with Plate fluorometer.
9. Resolve standard curve with Origin 2016 and logistic fit.

microcystin-LR (MC-LR) standard

MC-LR (Enzo Life sciences, ALX350-431)

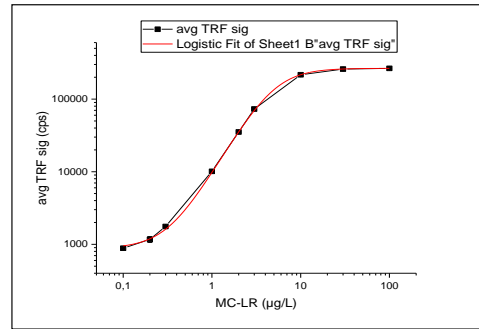
Prepared original stock of 1000 µg/L in reagent water+5%Methanol. Stored at (-20C)

Further working standard solution in reagent water: 100, 30, 10, 3, 2, 1, 0.3, 0.2 and 0.1 µg/L (stored at 4C for short term, -20 C for long term)

Reagent mixture in assay buffer

- 1 µg/mL biotinylated anti-ADDA Antibody; +
- 1 µg/mL anti-immunocomplex scFv-AP +
- 0.5 µg/mL NI-Eu-anti AP pAb.

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standard curve of microcystin-LR

(x)	TRF signal (counts per second)			(y)			blk+3SD (n=9)
	A	B	C	avg sig	std	cv%	
MC-LR (µg/L) std							
0	872	909	930				
0	914	864	886				
0	874	910	802	885	38	4.3	999
0.1	866	916	870	884	28	3.1	
0.2	1280	1170	1064	1171	108	9.2	
0.3	1786	1714	1779	1760	40	2.3	
1	10199	9647	10504	10117	434	4.3	
2	35200	35206	35297	35234	54	0.2	
3	73734	72210	72935	72960	762	1.0	
10	213098	217166	218035	216100	2636	1.2	
30	252353	264901	260336	259197	6351	2.5	
100	263641	265503	265481	264875	1069	0.4	

samp	Date	Dilution	TRF signal			(y)			*(x) From standard curve using logistic fit (origin)				
			A	B	C	Avg	sig	comments	std dev	cv%	conc µg/L	DF	1x conc (µg/L)
samp1	09.08.2019_1	(1x)	1272	1221	1080	1191	sig at lower range	99	8.4	0.20	1	0.20	
samp2	14.08.2019_2	(1x)	1092	1250	1050	1131	sig at lower range	105	9.3	0.18	1	0.18	
			detection limit			999							0.13

Interpretation

Samples were stored at -20°C until analysis, which was performed on 15.08.2019.

Before analysis, samples were heated at 80 °C for 10 min to release any possible cell bound toxins. Hence, the results represent the total peptide hepatotoxin amount (already released toxin in water and the cell bound toxin). The immunoassay (Akter et al., 2016) detects cyanobacterial peptide hepatotoxins (eg microcystins). For quantification, microcystin-LR was used as standard.

Sample 1: 09.08.2019 Littoistenjärvi hiekkaranta, knee dept water, ~15-20 cm below surface. Turbid water, weather: ~19°C.
Sample 2: 14.08.2019 Littoistenjärvi hiekkaranta, knee dept water, ~15-20 cm below surface. Clear water. weather: ~15°C

Result:

In Littoistenjärvi water, the detected cyanobacterial peptide hepatotoxin concentrations (free and cell bound) were as follows:

Sample 1 (09.08.2019, Littoistenjärvi hiekkaranta): ~ 0,2 µg/L

Sample 2 (14.08.2019, Littoistenjärvi hiekkaranta): below 0,2 µg/L